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Author-formatted, not peer-reviewed document posted on 23/09/2021

DOI: <https://doi.org/10.3897/arphapreprints.e75596>

**New and highest record of the rare Szechwan ratsnake,
Euprepiophis perlaceus (Serpentes: Colubridae) from
Muge Co, Kangding, Garzê, China**

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1 Article type: short communication

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3 **New and highest record of the rare Szechwan ratsnake,**
4 ***Euprepiophis perlaceus* (Serpentes: Colubridae) from Muge Co,**
5 **Kangding, Garzê, China**

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7 Dedicated to Klaus-Dieter Schulz (†2019)

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21 **Abstract**

22 Little is known about the distribution, biology, and ecology of the rare, endemic
23 Szechwan ratsnake. Here, molecular data of the species are provided from a habitat at 3370 m
24 a.s.l., the so far highest record of this snake. Genetic analysis (mtDNA and nDNA) confirmed
25 the identity of the specimen as *E. perlaceus* and its close relationship to *E. mandarinus*. P-
26 distances are in the range known for Elaphids and *Euprepiophis* as interspecific.

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28 Key words: *Euprepiophis*, high elevation, Muge Co lake basin, southeast margin of Tibet,
29 species identification

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31 **Introduction**

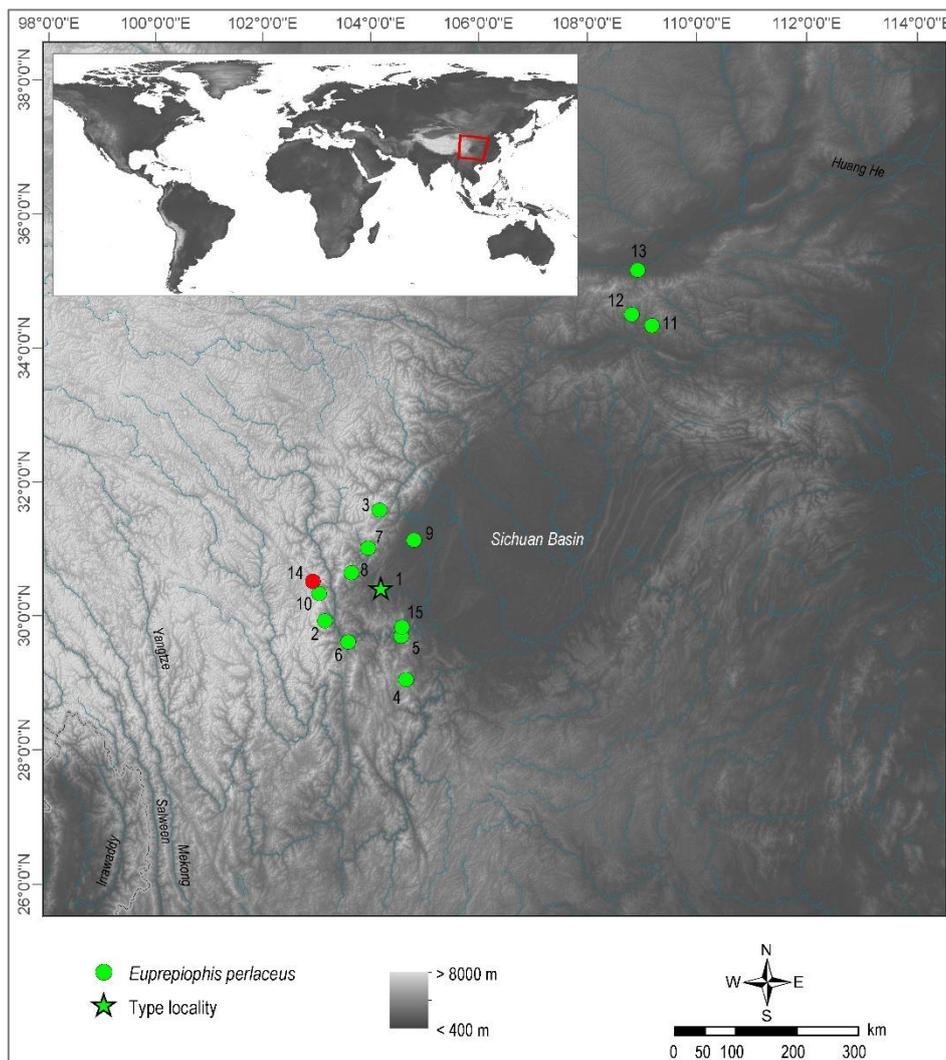
32 The Szechwan ratsnake, *Euprepiophis perlaceus* (Stejneger, 1929), had been
33 originally described as *Elaphe perlacea* based on a single male specimen from Yachow
34 (Ya'an Prefecture), Sichuan Province, China (holotype USNM 76257), Fig. 1. Schulz (1989)
35 suggested assigning this species as a subspecies of *Elaphe mandarinus* due to the
36 morphological similarity and overlapping distribution range of the two taxa. Morphological
37 data of three additional *Elaphe perlacea* specimens were presented by Zhao (1990). More
38 recently, the phylogenetic relationships within the genus *Elaphe* were revised, placing *Elaphe*
39 *mandarina* and *Elaphe conspicillata* in the genus *Euprepiophis*, as well as *Elaphe perlacea*
40 because of its close morphological relationship with these two species (Utiger et al. 2002).
41 However, the taxonomic status of the Szechwan ratsnake remained controversial until a
42 phylogenetic study which included three specimens of that species from unknown localities
43 (Chen et al. 2014). Based on genetic data and coalescent models, this study determined
44 *Euprepiophis perlaceus* as distinct species. Supporting morphological data for the species
45 validity were recently provided, based on 20 specimens collected in Shimian County,
46 Chongzhou City, and Kangding County (Zhou et al. 2019).

47 Very little is known about the ecology, biology, and distribution of *Euprepiophis*
48 *perlaceus*. So far, this species has been recorded from mountainous regions between
49 elevations of 1600 and 2400 m a.s.l., within a narrow range in western Sichuan, and Shaanxi
50 Province, China (Fig. 1; supplementary Tab. S1) where it is found in natural, humid broad-

51 leaf forests and shrubby grassland (Ding et al. 2017; Hu et al. 2002; Tang et al. 2021). Their
 52 life history characteristics and ecology probably resemble those of *E. mandarinus* (Schulz
 53 1996).

54 In the present study, molecular data, and a new record of *Euprepiophis perlaceus* from
 55 Sichuan are provided. Mitochondrial and nuclear DNA sequence data were used to validate
 56 the identity of the specimen by assigning them to existing *E. perlaceus* sequences. These
 57 recent data may support future research on this taxon in the mountains of Western Sichuan
 58 and potentially suitable adjacent regions.

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 61 **Figure 1.** Map showing the locality of the *Euprepiophis perlaceus* specimen reported herein
 62 (red dot; locality number 14) and further known records (for details, see supplementary Tab.
 63 S1). The type locality of *E. perlaceus* is indicated by a star.

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65 **Methods**

66 **Tissue sample, DNA sequencing and analysis**

67 Two adult individuals (one dead, one alive) were found in Muge Co area of Gongge
 68 mountains, Kangding County, Garzê, China (30.19°N, 101.87°E, 3370 m; Fig. 1,
 69 supplementary Tab. S1). A scale sample was taken from the dead specimen, transferred into
 70 absolute ethanol, and stored at -20 °C. Genomic DNA was extracted from tissue using the
 71 DNeasy Blood & Tissue Kit (Qiagen, Venlo, Netherlands) following the manufacturer's
 72 protocol. Approximately 351 bp of the 12S and 446 bp of the 16S ribosomal RNA (rRNA),
 73 666 bp of the COI, 662 bp of the ND2, 678 bp of the ND4, 781 bp of the cytb, and a fragment

74 of 567 bp of the nuclear *c-mos* gene were amplified via the polymerase chain reaction (PCR)
 75 using primers and PCR conditions as previously described (12S: Utiger et al. 2002; 16S:
 76 Hedges 1994; COI: Nagy et al. 2012; ND2: L4437b/tRNA-trpR, Macey et al. 1997, Ashton
 77 and de Queiroz 2001; ND4: Forstner et al. 1995; cytb: H15547/L14724, Edwards et al. 1991,
 78 Irwin et al. 1991; *c-mos*: Lawson et al. 2005). PCR products were purified using the mi-PCR
 79 Purification Kit (Metabion, Planegg, Germany) and sequenced in both directions by
 80 Macrogen (Amsterdam, Netherlands; <http://www.macrogen.com>).

81 We aligned the new sequences (accession numbers 12S: xxx, 16S: xxx, COI: xxx,
 82 ND2: xxx, ND4: xxx; cytb: xxx; *c-mos*: xxx) to data of the three *Euprepiophis* species,
 83 *Elaphe taeniura*, *Elaphe carinata*, and *Ptyas korros* (outgroup) available from GenBank
 84 (supplementary Tab. S1) by eye. Alignment of protein coding regions based on nucleotides
 85 and amino acids produced similar results, since no ambiguities, such as deletions, insertions,
 86 or stop codons, were found. The final concatenated rRNA + mtDNA + nDNA sequence
 87 dataset consisted of 11 taxa and contained 4151 alignment positions of which 590 were
 88 phylogenetically informative. We generated a Bayesian-inference (BI) tree using MrBayes
 89 v.3.2.6 (Ronquist et al. 2012). The dataset was partitioned a priori by gene and codon
 90 fragments, and PartitionFinder 1.1.1 (Lanfear et al. 2012) was applied to optimize partitions
 91 using linked branch lengths, the corrected Akaike Information Criterion, the greedy search
 92 algorithm, and the substitution models implemented in MrBayes. Analysis was run for 10
 93 million generations, with four parallel Markov chain Monte Carlo simulations and four chains
 94 (one cold and three heated), sampling trees every 1000th generation. Inspection of the
 95 standard deviation of split frequencies after the final run as well as the effective sample size
 96 value of the traces using Tracer v. 1.7.1 (Rambaut et al. 2018) indicated convergence of
 97 Markov chains. The first 25% of the samples of each run were dropped as burn-in.

98 Pairwise uncorrected p-distances were assessed separately for *c-mos* and ND2
 99 sequence data using Mega-X v.11.0 (Kumar et al. 2018) with the pairwise deletion option,
 100 1000 bootstrap replications, and by considering both transitions and transversions.

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102 Results and Discussion

103 The first Szechwan ratsnake was spotted near a road surrounded by deciduous broad-
 104 leaved forest, moving fast under rocks. The second snake was found dead beside the road, not
 105 far from the first individual (Figs. 2).

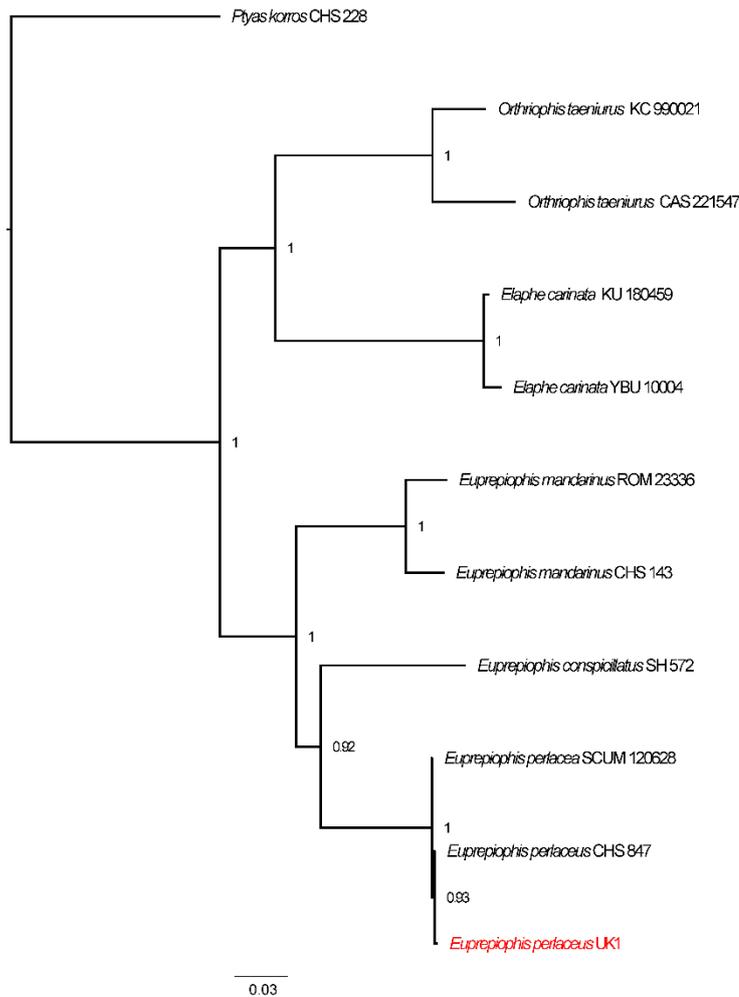
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107 **Figure 2.** Dead specimen of *Euprepiophis perlaceus* (left panel) found in a mountainous
 108 habitat (right panel) at an elevation of 3370 m a.s.l. in Muge Co area, Kangding County,
 109 Garzê, China.

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The identity of the dead snake specimen was confirmed as *Euprepiophis perlaceus*. Our sequences nested in the clade of *E. perlaceus*; the placement within this clade was highly supported (Fig. 3). Pairwise p-distances between mitochondrial ND2 sequences of different *Euprepiophis* species ranged between 11.9 (*E. conspicillatus*/*E. perlaceus*) and 14.2 % (*E. conspicillatus*/*E. mandarinus*); for c-mos p-distances were 0.53 (*E. conspicillatus*/*E. mandarinus*; *E. mandarinus*/*E. perlaceus*) and 0.71 % (*E. conspicillatus*/*E. perlaceus*), respectively (Tab. 1, 2). These genetic distances are moderately lower than the evolutionary distances between *Elaphe* species but in the range of interspecific distances.



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Figure 3. Bayesian inference tree based on concatenated rRNA, mtDNA and nDNA sequence data inferred with MrBayes. Species name is followed by the sample or voucher ID. Numbers on branch nodes represent posterior probability values. ID of the sample sequenced herein is indicated red.

Table 1. Pairwise distance matrix (lower: uncorrected p-distances, %; upper: standard error) for ND2 sequences of samples. Cells with intraspecific distances are indicated grey.

	<i>carinata</i>	<i>carinata</i>	<i>taeniurus</i>	<i>taeniurus</i>	<i>conspicillatus</i>	<i>mandarinus</i>	<i>mandarinus</i>	<i>perlaceus</i>	<i>perlaceus</i>	UK1
<i>E. carinata</i> KU180459		0.40	2.22	2.39	2.46	2.52	2.47	2.26	2.26	2.34
<i>E. carinata</i> YBU10004	1.06		2.24	2.42	2.52	2.60	2.55	2.33	2.33	2.40
<i>E. taeniura</i> AH015912	18.00	18.40		1.19	2.54	2.46	2.45	2.30	2.30	2.29
<i>E. taeniurus</i> KC990021	19.75	20.16	7.05		2.58	2.44	2.42	2.25	2.25	2.35

<i>E. conspicillatus</i> DQ902213	19.77	20.38	20.37	20.38		1.87	1.90	1.61	1.61	1.66
<i>E. mandarinus</i> CHS143	19.68	20.29	19.60	19.48	14.12		1.00	1.76	1.76	1.87
<i>E. mandarinus</i> ROM23336	19.95	20.57	19.86	19.27	14.24	5.22		1.73	1.73	1.82
<i>E. perlaceus</i> CHS847	18.10	18.70	17.59	17.52	11.85	12.55	12.21		0.00	0.00
<i>E. perlaceus</i> SCUM120628	18.10	18.70	17.59	17.52	11.85	12.55	12.21	0.00		0.00
UK1 (this study)	18.35	18.78	17.01	17.63	11.71	13.05	12.49	0.00	0.00	

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Table 1. Pairwise distance matrix (lower: uncorrected p-distances, %; upper: standard error) for c-mos sequences of samples.

	<i>carinata</i>	<i>conspicillatus</i>	<i>mandarinus</i>	UK1	<i>taeniura</i>
<i>E. carinata</i> YBU10004		0.47	0.46	0.54	0.35
<i>E. conspicillatus</i> DQ902065	1.28		0.30	0.36	0.38
<i>E. mandarinus</i> ROM23336	1.28	0.53		0.29	0.42
UK1 (this study)	1.65	0.71	0.53		0.46
<i>E. taeniura</i> CAS221547	0.73	0.89	1.07	1.25	

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The new record matches the preferred habitat conditions of *E. perlaceus*, which include deciduous broadleaf forest and mixed evergreen deciduous broad-leaved forest in temperate mountains (Ding et al. 2017; Gan et al. 2017; Tang et al. 2021). However, given the elevation of the new observation, the vertical distribution of the snake seems to be wider than previously reported. Thus, potential habitats of the species might be available in large parts of the Hengduan Shan and mountains around the Sichuan Basin, e.g., the Daba Shan, Qinling Shan and even the Wu Shan. Indeed, a recent study reported on records of *E. perlacea* from Qinling Mountains, Foping County, Shaanxi Province and suspected a continuous distribution range of the snake in the neighboring provinces Sichuan and Gansu (Tang et al. 2021).

According to the on IUCN Red List assessments of Asian snake species (DECISION 16.104), major threats for the Szechwan ratsnake are habitat loss, and overharvesting for food and skins. Another threat is the increasing, unregulated trade in this snake for pet (Tang et al. 2021), especially in the European and American markets, highlighting the need for detailed *in situ* research (e.g., biology, distribution).

Conclusions

Here a new and so far, highest record of *Euprepiophis perlaceus* from Kangding County is reported. Species validation is based on multi-loci DNA sequence data of *E. perlaceus* and congeners. The study provides the first high-altitude record of the Szechwan ratsnake, supporting the assumption of a wider distribution of that species than previously believed.

Acknowledgements

This work was funded by the German Research Foundation (DFG 271/21).

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